

Intestinal cholesterol secretion: future clinical implications

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ABSTRACT

Together with the liver, the intestine serves as a homeostatic organ in cholesterol metabolism. Recent evidence has substantiated the pivotal role of the intestine in reverse cholesterol transport (RCT). RCT is a fundamental antiatherogenic pathway, mediating the removal of cholesterol from tissues in the body to the faeces. In humans, faecal cholesterol elimination via the RCT pathway is considered to be restricted to excretion via the hepatobiliary route. Recently, however, direct trans-intestinal excretion of plasma-derived cholesterol (TICE) was shown to contribute substantially to faecal neutral sterol (FNS) excretion in mice. TICE was found to be amenable to stimulation by various pharmacological and dietary interventions in mice, offering new options to target the intestine as an inducible, cholesterol-excretory organ. The relevance of TICE for cholesterol elimination in humans remains to be established. There is, however, emerging evidence for the presence of TICE in human (patho) physiology. This review discusses our current understanding of TICE and its novel therapeutic potential for individuals at increased risk of cardiovascular disease.

KEYWORDS

Reverse cholesterol transport, intestine, cardiovascular disease

INTRODUCTION

In the human body, cholesterol homeostasis is tightly regulated. This is not only of physiological importance, but also bears clinical relevance, since excessive cholesterol accumulation in the arterial wall invariably leads to the development of atherosclerotic cardiovascular disease

(CVD). Although the inhibition of cholesterol synthesis by statins has resulted in a powerful reduction of CVD risk with a mean relative reduction of 25%,¹ there is still an unmet need for additional effective therapies to further reduce the residual CVD risk. In the past decade, research has mainly focussed on high-density lipoprotein cholesterol (HDL-C) raising therapies,^{2,3} because of the strong inverse relationship between plasma HDL-C concentrations and CVD risk in epidemiological studies.^{4,7} However, recent studies, aimed at increasing plasma HDL-C concentrations, have not substantiated a significant CVD risk reduction.⁷⁻¹⁰ Hence, rather than aiming for an increase of HDL-C concentration, current research is focussed on elucidating and, if feasible, quantifying the mechanisms contributing to the atheroprotective properties of HDL-C.

The most established protective function of HDL-C is its role in the reverse cholesterol transport (RCT). This process was originally defined as the efflux of cholesterol from peripheral tissues, including arterial intra-plaque macrophages, subsequent transport in the plasma and uptake by the liver, followed by biliary secretion and elimination via the faeces.¹¹ Faecal excretion is the predominant way for eliminating cholesterol, because apart from conversion to bile acids, cholesterol cannot be catabolised to a significant extent within the human body. The classical RCT concept rests on two principles: 1. HDL-C is the primary lipoprotein involved in RCT and 2. biliary secretion is the sole route for intestinal elimination of plasma-derived cholesterol. In view of recent findings, both of these principles need to be reconsidered. The first is beyond the scope of this review. In short, in contrast to the current consensus, several studies have shown that plasma HDL-C levels do not determine biliary or faecal excretion of cholesterol in mice,¹²⁻¹⁴ whereas

studies in humans have yielded conflicting results.¹⁵⁻¹⁸ This review handles the second paradigm, the obligatory role of hepatobiliary cholesterol secretion in RCT. This historical concept has recently been challenged by studies in mice, indicating the existence of direct trans-intestinal cholesterol excretion (TICE) as an alternative cholesterol-eliminating pathway.

TRANS-INTESTINAL CHOLESTEROL EXCRETION: ANIMAL STUDIES

Cholesterol destined for hepatobiliary cholesterol secretion is taken up at the basolateral side of the hepatocyte via a number of lipoprotein receptors and is subsequently secreted at the canalicular membrane by a not fully elucidated secretion process, mediated for the largest part by the ATP-binding cassette G5/G8 (*abcg5/g8*) transporter.¹⁹ If hepatobiliary cholesterol secretion were the primary route for cholesterol elimination, then inhibition of *abcg5/g8* could be expected to result in extreme reductions of faecal neutral sterol (FNS) excretion. Interestingly, *abcg5* and *abcg8* double-knockout mice did not show these expected reductions in FNS loss.^{20,21} Similar observations were made in other murine models of impaired hepatobiliary cholesterol secretion.²²⁻²⁵ These studies unambiguously point towards the existence of an alternative, non-biliary cholesterol excretion pathway, at least in mice with genetically hampered hepatobiliary cholesterol secretion.

The concept of a non-biliary cholesterol excretion route is not novel. Already in 1927, it was demonstrated that FNS loss was paradoxically increased in dogs undergoing surgical bile diversion, as compared with control dogs.²⁶ These early findings were confirmed in a replication study in 1973,²⁷ as well as in studies in bile-diverted rats.^{28,29} Similar observations have been made in the human situation, as described below.

More recently, additional murine intestinal perfusion studies and *in vivo* stable isotope studies substantiated that this alternative TICE route is also present in mice with intact hepatobiliary secretion and enterohepatic cycling.^{30,31} In these studies, TICE accounted for roughly 20-33% of FNS loss. Moreover, the intestinal perfusion studies showed that plasma cholesterol can directly traverse the small intestine in a basolateral to apical direction, stimulated by the luminal presence of bile salt and phospholipid acceptors.³² Furthermore, TICE was found to occur predominantly in the proximal part of the small intestine.^{25,30} Importantly, a recent study showed that faecal excretion of macrophage-derived cholesterol can also proceed in the absence of biliary sterol secretion, suggesting that TICE can also mediate reverse cholesterol transport from cholesterol-loaded macrophages.³³ This implies that TICE may have

antiatherogenic effects. However, a similar study could not confirm these results, for as yet unknown reasons.³⁴ Hence, it remains to be established whether induction of cholesterol elimination via TICE results in inhibition of atherosclerosis progression.

The molecular mechanisms underlying TICE are not fully understood. Hence, it is not known whether TICE is an active, transporter-mediated metabolic process. In order to effectively target this pathway, the following items need to be addressed: characterisation of plasma donor particles delivering cholesterol to the intestine for subsequent excretion via TICE; identification of transporters located at the basolateral membrane of intestinal cells, involved in the uptake of cholesterol destined for intestinal excretion; elucidation of intracellular trafficking mechanisms by which cholesterol is transported towards the apical membrane of enterocytes; identification of all apically located transporters and potential luminal acceptors which facilitate the excretion of cholesterol to the enteric lumen.

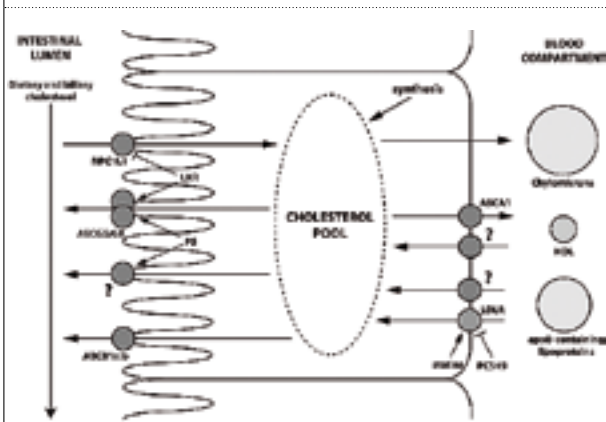
Thus far, some progress has been made, which has been the focus of recent comprehensive reviews.³⁵⁻³⁷ The most important findings are summarised below (see also *figure 1*).

Despite its classical role in RCT, several studies indicate that plasma HDL is not the donor particle delivering plasma cholesterol for elimination via TICE. *Abca1* and *apoA1*-deficient mice, expressing negligible plasma HDL-C concentrations, exhibit normal or increased FNS excretion under normal conditions¹²⁻¹⁴ and intestinal secretion of radiolabelled plasma-derived cholesterol was unaltered in *abca1*^{-/-} mice as compared with their wild-type littermates.³⁸

Instead, a recent review provides evidence to show that very-low-density lipoprotein (VLDL) remnants or further catabolic products of VLDL may serve as plasma donor particles delivering cholesterol for TICE.³⁵ In line, previous kinetic studies established a relatively high uptake rate of LDL cholesterol (LDL-C) into the intestine.³⁹ The LDL receptor (LDL-R) or one of the other receptors in the LDL-R family have been proposed as the basolateral transporters that mediate TICE.^{35,40} However, experiments in *ldl-r*^{-/-} mice failed to substantiate this concept.^{25,35} Further studies are required to elucidate the intracellular itinerary of cholesterol destined for excretion via TICE.

The *abcg5/g8* transporter, located at the brush border membrane of the small intestine, is likely to facilitate the last step of TICE. This is supported by several murine studies using various methodologies to quantify TICE.^{31,41-43} In contrast, *abcg5/g8* function was not found to affect TICE as measured in intestinal perfusion,³⁰ most likely reflecting methodological differences. Hence, TICE cannot be fully attributed to the activity of *abcg5/g8*, as a significant amount of TICE is still present in *abcg5* or *abcg8*-deficient

Figure 1. Schematic representation of cholesterol fluxes in enterocytes related to the TICE pathway. The cholesterol pool of the intestinal cell is fuelled by uptake from the intestinal lumen via apically localised NPC1L1, endogenous synthesis, and via uptake of cholesterol from HDL and apoB-containing lipoproteins on the basolateral side (blood compartment). Apically, the main contributor to TICE is the ABCG5/G8 heterodimer; ABCB1a/b might also play a role as well as an additional route that has not yet been identified. On the basolateral side, the main cholesterol donors for TICE seem to be apoB-containing lipoproteins in a pathway that is likely to involve modulation of LDL-R expression. TICE can be increased by LXR activation as well as dietary plant sterols, partly dependent on functional ABCG5/G8 expression. The role of different intracellular pathways of cholesterol trafficking connecting basolateral uptake and apical secretion is currently unclear



ABC = ATP-binding cassette; apoB = apolipoprotein B; HDL = high-density lipoprotein; LDL-R = low-density lipoprotein receptor; LXR = liver X receptor; NPC1L1 = Niemann-Pick C1 Like 1; PCSK9 = pro-protein convertase subtilisin kexin type 9; PS = plant sterols; TICE = transintestinal cholesterol excretion. (Adapted from: Tietge UW, Groen AK. Role of the TICE? Advancing the concept of transintestinal cholesterol excretion. *Arterioscler Thromb Vasc Biol.* 2013;33:1452-3).

mice. Other apically located proteins are likely to be involved. In fact, a recent report suggests that the *abcbla/b* protein may serve as an apical excretory transporter in TICE.⁴⁰ Finally, it is plausible that acceptors in the intestinal lumen are required for cholesterol excretion via TICE. Bile acids and phospholipids in the intestinal lumen have been shown to stimulate the amount of cholesterol excreted via TICE.^{30,32} The dependence on phospholipids is analogous to hepatic cholesterol secretion into the bile.²³ In the absence of these acceptors, only a small mass of TICE could be observed, which was probably attributable to shedding of enterocytes. Finally, the Niemann-Pick C2 (NPC2) protein was recently found to stimulate *abcg5/g8*-dependent biliary cholesterol secretion without affecting the *abcg5/g8*-independent pathway.^{44,45} Although

speculative, NPC2 might function as an acceptor for TICE mediated by *abcg5/g8*. Additional studies focusing on these acceptors might yield therapeutic interventions that would not require systemic distribution.

TRANS-INTESTINAL CHOLESTEROL EXCRETION: HUMAN STUDIES

The extent to which TICE contributes to faecal cholesterol elimination in humans remains to be established. Until recently, indications of the presence of TICE in human physiology were predominantly based on studies in patients with bile fistulae. In patients with complete biliary obstruction, a substantial portion of faecal sterols was found to be of non-dietary origin⁴⁶ and in another study in bile-diverted patients, the intestinal mucosa was found to secrete 250-400 mg of cholesterol per day.⁴⁷ A human intestinal perfusion study corroborated the presence of TICE, showing that approximately 44% of total FNS output originated from non-biliary origin.⁴⁸ These and a number of other reports⁴⁹⁻⁵¹ were mostly disregarded, likely pertaining to the small series of observations and the study limitations of the bile-diversion conditions. These include hampered cholesterol absorption and strongly upregulated cholesterol and bile acid synthesis. Furthermore, limitations of intestinal perfusion studies may have contributed, such as the absence of food, biliary and pancreatic components in the rinsed and perfused intestinal segments, together with the specific composition of the perfusate, which may have influenced the excretory capacity of enterocytes. Hence, studies on the non-biliary cholesterol excretion route were not pursued, until the more recent animal studies described above. Moreover, recent *in vitro* experiments using jejunal explants from humans showed activity of the TICE pathway for the first time in humans.⁴⁰ In these experiments, TICE depended on the presence of oxygen and was significantly decreased at low temperatures, which suggests that TICE is an active metabolic process.

Although the currently available human data collectively lend support to the presence of TICE in human cholesterol metabolism, a definite answer to this question has remained elusive. This is largely due to the technical challenges faced to reliably estimate this flux in humans *in vivo*, which requires simultaneous assessment of cholesterol absorption, biliary secretion and FNS excretion. We have recently attempted to quantify TICE in a population of mildly hypercholesterolaemic humans, by combining our previous experience from validated stable cholesterol isotope methodologies in mice³¹ and humans.^{15,52-53} Our unpublished data indicate that TICE is indeed present in human physiology and that it is sensitive to pharmacological stimulation, as described below.

TRANS-INTESTINAL CHOLESTEROL EXCRETION: FUTURE THERAPEUTIC POTENTIAL

The TICE pathway was found to be sensitive to various forms of dietary and pharmacological activation. However, at present, this is mostly confined to preclinical studies.

Liver-X-receptor agonists

Liver X nuclear receptors (LXRs) play a central role in cholesterol metabolism. Upon activation, LXRs induce expression of a series of genes that are involved in cholesterol efflux, absorption, transport and excretion.⁵⁴ Consistently, LXRs limit the development of atherosclerosis in mice and are therefore considered promising therapeutic targets for CVD risk reduction.⁵⁵ However, activation of LXRs concurrently promotes hepatic *de novo* lipogenesis, steatosis, and hypertriglyceridemia via direct activation of the sterol regulatory element-binding protein-1c (*SREBP-1c*) gene and fatty acid synthesis pathways.⁵⁶ LXR agonists were found to stimulate TICE up to sixfold in murine studies using different experimental methodologies.^{23,30,31}

Recently, intestine-specific LXR agonists, which evade the unfavourable LXR-mediated effects on hepatic lipogenesis, have been developed. Studies indicate that intestine-specific activation of LXR, either genetic⁵⁷ or pharmacological,⁵⁸ is crucial for LXR-induced atheroprotection. Although it is tempting to suggest that TICE underlies parts of these favourable sequelae, a study which directly shows that TICE is stimulated by intestine-specific LXR agonists has not yet been reported. Finally, although promising in animal studies, the development of LXR-targeted drugs has largely been discontinued due to observations of marked increases in plasma apoB containing lipoproteins and/or a marked liver-steatotic response. To the best of our knowledge, there are no ongoing trials with intestine-specific LXR agonists. Hence, clinical studies evaluating their effects on TICE and atherosclerosis are not expected in the very near future.

Ezetimibe

Ezetimibe inhibits intestinal cholesterol absorption⁵⁹ in both mice and men, accomplished through inhibition of the Niemann-Pick C1 Like 1 (NPC1L1) transporter.⁶⁰ Despite a compensatory increase in endogenous cholesterol biosynthesis,⁶⁰ ezetimibe monotherapy lowers plasma LDL-C concentrations by approximately 15-20%.⁵⁹ Ezetimibe has been shown to stimulate RCT from macrophages in mice, via as yet unidentified mechanisms.^{61,62} Furthermore, when assessing cholesterol balance in ezetimibe-treated mice, the enhancement in FNS excretion cannot be attributed to cholesterol absorption inhibition or increased biliary cholesterol

secretion alone.⁴¹ In line, it has been suggested that ezetimibe might stimulate FNS excretion through stimulation of TICE,⁶³ although this was contradicted by another murine intestinal perfusion study.⁶⁴ Yet, our unpublished results of *in vivo* stable isotope studies in both mice and men showed a striking effect of ezetimibe on TICE [unpublished results, Jakulj, Stroes, Groen].

The underlying mechanisms by which ezetimibe might stimulate TICE are unknown. We speculated that the inhibition of NPC1L1 disturbs normal intracellular vesicle trafficking leading to increased transport of cholesterol to the apical membrane of the enterocytes.⁴¹ Another possibility is that ezetimibe exerts its stimulatory effect on TICE by manipulation of the intraluminal bile acid and phospholipid content.^{32,65,66}

Although our findings suggest an alternative mode by which ezetimibe might reduce plasma cholesterol concentrations and possibly reduce CVD risk, the latter issue is still precarious. Despite preclinical evidence that ezetimibe is atheroprotective,⁶⁷ to date, clinical studies have not been able to substantiate this: ezetimibe failed to regress carotid intima media thickness (cIMT) progression in patients with familial hypercholesterolaemia in the ENHANCE trial⁶⁸ and was found to be inferior to niacin in patients with coronary heart disease in the ARBITER-6 HALTS trial.⁶⁹ Next to major methodological disadvantages,^{68,70} several off-target effects,⁷¹ as well as upregulation of HMG-CoA reductase expression,⁷² have been proposed as potential explanations. However, in the ARBITER-6 HALTS study, ezetimibe did hamper cIMT progression in statin-treated patients with fairly low LDL-C concentrations, who would thereby not likely to be considered for ezetimibe add-on therapy.⁶⁹ Furthermore, not all cIMT trials investigating ezetimibe have been negative.⁷³ A large clinical study of 18,000 patients, the IMPROVE-IT trial, is underway to determine whether additional cholesterol lowering by ezetimibe on top of statins can be translated into a reduction in cardiovascular event rate.⁷⁴ Although this trial started in 2005 and the results were expected in 2011, outcomes are still awaited, supposedly due to recruitment of additional patients after an unfavourable interim analysis. This trial is conducted in patients who have suffered from an acute coronary syndrome and who expressed low LDL-C concentrations at baseline, as inclusion of patients with higher LDL-C concentrations would not have achieved guideline-recommended LDL-C concentrations under the trial protocol, which would have been ethically unacceptable. Hence, it is conceivable that no additional benefit can be gained in this population, if ezetimibe's effect on atherosclerosis is causally related to plasma LDL-C reductions alone. Release of the study outcomes has been postponed until September 2014 (ClinicalTrials.gov:NCT00202878).

PCSK9

Proprotein convertase subtilisin kexin type 9 (PCSK9) is a secreted protein that reduces the amount of LDL-R at the cell surface of primarily the liver. PCSK9 circulates in the blood and binds the extracellular domain of the LDL-R to produce post-translational down-regulation of this receptor in lysosomes.⁷⁵ Loss-of-function mutations in the *PCSK9* gene result in 15-18% reductions in plasma LDL-C concentrations and carriers of these mutations express a 47-88% reduction in CVD risk.⁷⁶

Next to the liver, PCSK9 is abundantly expressed in the intestine and it has been shown that PCSK9 modulates cholesterol transport and metabolism, as well as production of apoB-containing lipoproteins, in intestinal cells.⁷⁷ A recent report by the same research group revealed that PCSK9 is a repressor of TICE and that acute repression depended on a functional LDL-R.⁴⁰

These findings may be of clinical importance, as Phase I and II PCSK9-inhibiting treatment modalities, such as single-stranded antisense DNA-like oligonucleotides or double-stranded small interference RNA, have shown promising results in terms of LDL-C lowering.⁷⁵ Phase III trials with longer duration and larger patient populations are currently underway, which should also establish whether PCSK9 inhibition reduces cardiovascular event rate in humans.

Plant sterols

Plant sterols are not endogenously synthesised by humans, but are strictly derived from the diet. They perform functions in plant cells similar to those of cholesterol in mammalian cells. Campesterol and sitosterol are the most abundant ones. They share a high degree of structural similarity with cholesterol, but are much more hydrophobic. Plant sterols are present in small amounts of fruits, vegetables, nuts, seeds and edible oils; marketed sources are primarily derived from soybean and pine tree oil. Total dietary plant sterol consumption in the average Western diet is 150-350 mg per day.^{78,79} Daily consumption of 2g of plant sterols is associated with LDL-C reductions varying from 4-15% in hypercholesterolaemic or normocholesterolaemic adults.⁸⁰

Plant sterols are thought to displace cholesterol from incorporation into micelles, thereby limiting cholesterol absorption in the intestinal lumen by approximately 25-36%.⁸¹ However, additional cholesterol-lowering mechanisms have been postulated.⁸² Interestingly, the cholesterol-lowering effects of plant sterol consumption were recently ascribed to stimulation of intestinal cholesterol excretion via TICE, as plant sterol feeding resulted in a sixfold induction of TICE in wild-type mice.⁴³ This is supported by a recent crossover plant sterol feeding trial in 18 adults, who in random order consumed dietary plant sterols from negligible (0 mg) to high (2g)

amounts, resulting in a dose-dependent increase in FNS output, which could not be explained by the corresponding reductions in measured cholesterol absorption.⁸³

The mechanisms by which plant sterols could stimulate TICE are currently unknown. Several mechanisms have been suggested, including both LXR-dependent and -independent mechanisms.⁸² It is less likely that the stimulation of TICE is LXR-mediated, as plant sterols did not alter LXR target genes in the study by Brufau *et al.*⁴³ and studies investigating plant sterols as possible ligands of LXR have been conflicting.^{21,84,85} A possible LXR-independent mechanism might include interference of plant sterols with cholesterol trafficking within the enterocyte, as plant sterols have been shown to affect expression of genes encoding proteins of the annexin family, which are involved in the regulation of membrane properties.⁸⁶ Besides studies aiming to unravel the underlying molecular mechanisms, human studies to assess the effect of plant sterols on TICE are also lacking at present.

CONCLUSIONS

In conclusion, trans-intestinal cholesterol excretion might serve as an attractive future target for LDL-C lowering and CVD reduction, provided underlying molecular mechanisms are elucidated. Although promising, the therapeutic potential of targeting the TICE pathway is to date confined to preclinical studies and it is unknown whether pharmacological targeting of the TICE pathway will also yield a clinical benefit. Available interventions that have been shown to stimulate TICE and may therefore warrant further clinical evaluation include ezetimibe, PCSK9-inhibitors and plant sterols.

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REFERENCES

1. Baigent C, Blackwell L, Emberson J, et al. Efficacy and safety of more intensive lowering of LDL cholesterol: a meta-analysis of data from 170,000 participants in 26 randomised trials. *Lancet*. 2010;376:1670-81.
2. Wright RS. Recent clinical trials evaluating benefit of drug therapy for modification of HDL cholesterol. *Cur Opin Cardiol*. 2013;28:389-98.
3. Bosch N, Frishman WH. Newer Therapeutic Strategies to Alter HDL Level and Function. *Cardiol Rev*. 2013 May 23.
4. Gordon DJ, Probstfield JL, Garrison RJ, et al. High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation*. 1989;79:8-15.

5. Assmann G, Schulte H, von Eckardstein A, Huang Y. High-density lipoprotein cholesterol as a predictor of coronary heart disease risk. The PROCAM experience and pathophysiological implications for reverse cholesterol transport. *Atherosclerosis*. 1996;124(Suppl):S11-20.
6. deGoma EM, DeGoma RL, Rader DJ. Beyond high-density lipoprotein cholesterol levels evaluating high-density lipoprotein function as influenced by novel therapeutic approaches. *J Am Coll Cardiol*. 2008;51:2199-211.
7. Barter PJ, Caulfield M, Eriksson M, et al. Effects of torcetrapib in patients at high risk for coronary events. *New Engl J Med*. 2007;357:2109-22.
8. Briel M, Ferreira-Gonzalez I, You JJ, et al. Association between change in high density lipoprotein cholesterol and cardiovascular disease morbidity and mortality: systematic review and meta-regression analysis. *BMJ*. 2009;338:b92.
9. Boden WE, Probstfield JL, Anderson T, et al. Niacin in Patients with Low HDL Cholesterol Levels Receiving Intensive Statin Therapy. *New Engl J Med*. 2011;365:2255-67.
10. Schwartz GG, Olsson AG, Abt M, et al. Effects of dalcetrapib in patients with a recent acute coronary syndrome. *New Engl J Med*. 2012;367:2089-99.
11. Glomset JA. The plasma lecithins:cholesterol acyltransferase reaction. *J Lipid Res*. 1968;9:155-67.
12. Groen AK, Bloks VW, Bandsma RH, Ottenhoff R, Chimini G, Kuipers F. Hepatobiliary cholesterol transport is not impaired in Abca1-null mice lacking HDL. *J Clin Invest*. 2001;108:843-50.
13. Jolley CD, Woollett LA, Turley SD, Dietschy JM. Centripetal cholesterol flux to the liver is dictated by events in the peripheral organs and not by the plasma high density lipoprotein or apolipoprotein A-I concentration. *J Lipid Res*. 1998;39:2143-9.
14. Xie C, Turley SD, Dietschy JM. ABCA1 plays no role in the centripetal movement of cholesterol from peripheral tissues to the liver and intestine in the mouse. *J Lipid Res*. 2009;50:1316-29.
15. Harchaoui K El, Franssen R, Hovingh GK, et al. Reduced fecal sterol excretion in subjects with familial hypoalphalipoproteinemia. *Atherosclerosis*. 2009;207:614-6.
16. Eriksson M, Carlson LA, Miettinen TA, Angelin B. Stimulation of fecal steroid excretion after infusion of recombinant proapolipoprotein A-I. Potential reverse cholesterol transport in humans. *Circulation*. 1999;100:594-8.
17. Nanjee MN, Cooke CJ, Garvin R, et al. Intravenous apoA-I/lecithin discs increase pre-beta-HDL concentration in tissue fluid and stimulate reverse cholesterol transport in humans. *J Lipid Res*. 2001;42:1586-93.
18. Holleboom AG, Jakulj L, Franssen R, et al. In vivo tissue cholesterol efflux is reduced in carriers of a mutation in APOA1. *J Lipid Res*. 2013;54:1964-71.
19. Berge KE, Tian H, Graf GA, et al. Accumulation of dietary cholesterol in sitosterolemia caused by mutations in adjacent ABC transporters. *Science*. 2000;290:1771-5.
20. Yu L, Hammer RE, Li-Hawkins J, et al. Disruption of Abcg5 and Abcg8 in mice reveals their crucial role in biliary cholesterol secretion. *Proc Natl Acad Sci U S A*. 2002;99:16237-42.
21. Plösch T, Kok T, Bloks VW, et al. Increased hepatobiliary and fecal cholesterol excretion upon activation of the liver X receptor is independent of ABCA1. *J Biol Chem*. 2002;277:33870-7.
22. Temel RE, Tang W, Ma Y, et al. Hepatic Niemann-Pick C1-like 1 regulates biliary cholesterol concentration and is a target of ezetimibe. *J Clin Invest*. 2007;117:1968-78.
23. Kruit JK, Plösch T, Havinga R, et al. Increased fecal neutral sterol loss upon liver X receptor activation is independent of biliary sterol secretion in mice. *Gastroenterology*. 2005;128:147-56.
24. Schwarz M, Russell DW, Dietschy JM, Turley SD. Marked reduction in bile acid synthesis in cholesterol 7alpha-hydroxylase-deficient mice does not lead to diminished tissue cholesterol turnover or to hypercholesterolemia. *J Lipid Res*. 1998;39:1833-43.
25. Brown JM, Bell TA, Alger HM, et al. Targeted depletion of hepatic ACAT2-driven cholesterol esterification reveals a non-biliary route for fecal neutral sterol loss. *J Biol Chem*. 2008;283:10522-34.
26. Sperry W. Lipid Excretion IV. A study of the relationship of the bile to the fecal lipids with special reference to certain problems of sterol metabolism. *J Biol Chem*. 1927;71:351-78.
27. Pertsemilidis D, Kirchman EH, Ahrens EH. Regulation of cholesterol metabolism in the dog. I. Effects of complete bile diversion and of cholesterol feeding on absorption, synthesis, accumulation, and excretion rates measured during life. *J Clin Invest*. 1973;52:2353-67.
28. Dietschy JM, Siperstein MD. Cholesterol synthesis by the gastrointestinal tract: localization and mechanisms of control. *J Clin Invest*. 1965;44:1311-27.
29. Bandsma RH, Stellaard F, Vonk RJ, et al. Contribution of newly synthesized cholesterol to rat plasma and bile determined by mass isotopomer distribution analysis: bile-salt flux promotes secretion of newly synthesized cholesterol into bile. *Biochem J*. 1998;329(Pt 3):699-703.
30. Van der Velde AE, Vrins CLJ, van den Oever K, et al. Direct intestinal cholesterol secretion contributes significantly to total fecal neutral sterol excretion in mice. *Gastroenterology*. 2007;133:967-75.
31. Van der Veen JN, van Dijk TH, Vrins CLJ, et al. Activation of the liver X receptor stimulates trans-intestinal excretion of plasma cholesterol. *J Biol Chem*. 2009;284:19211-9.
32. Van der Velde AE, Vrins CLJ, van den Oever K, et al. Regulation of direct transintestinal cholesterol excretion in mice. *Am J Physiol Gastrointest Liver Physiol*. 2008;295:G203-G208.
33. Temel RE, Sawyer JK, Yu L, et al. Biliary sterol secretion is not required for macrophage reverse cholesterol transport. *Cell Metab*. 2010;12:96-102.
34. Nijstad N, Gautier T, Briand F, Rader DJ, Tietge UJF. Biliary sterol secretion is required for functional in vivo reverse cholesterol transport in mice. *Gastroenterology*. 2011;140:1043-51.
35. Temel RE, Brown JM. Biliary and nonbiliary contributions to reverse cholesterol transport. *Curr Opin Lipidol*. 2012;23:85-90.
36. Brufau G, Groen AK, Kuipers F. Reverse cholesterol transport revisited: contribution of biliary versus intestinal cholesterol excretion. *Arterioscler Thromb Vasc Biol*. 2011;31:1726-33.
37. Van der Velde AE, Brufau G, Groen AK. Transintestinal cholesterol efflux. *Curr Opin Lipidol*. 2010;21:167-71.
38. Vrins CLJ, Ottenhoff R, van den Oever K, et al. Trans-intestinal cholesterol efflux is not mediated through high density lipoprotein. *J Lipid Res*. 2012;53:2017-23.
39. Stange EF, Dietschy JM. Cholesterol synthesis and low density lipoprotein uptake are regulated independently in rat small intestinal epithelium. *Proc Natl Acad Sci U S A*. 1983;80:5739-43.
40. Le May C, Berger JM, Lespine A, et al. Transintestinal cholesterol excretion is an active metabolic process modulated by PCSK9 and statin involving ABCB1. *Arterioscler Thromb Vasc Biol*. 2013;33:1484-93.
41. Jakulj L, Vissers MN, van Roomen CP, et al. Ezetimibe stimulates faecal neutral sterol excretion depending on abcg8 function in mice. *FEBS Lett*. 2010;584:3625-8.
42. Yu L, York J, von Bergmann K, Lutjohann D, Cohen JC, Hobbs HH. Stimulation of cholesterol excretion by the liver X receptor agonist requires ATP-binding cassette transporters G5 and G8. *J Biol Chem*. 2003;278:15565-70.
43. Brufau G, Kuipers F, Lin Y, Trautwein EA, Groen AK. A reappraisal of the mechanism by which plant sterols promote neutral sterol loss in mice. *PLoS One*. 2011;6:e21576.
44. Yamanashi Y, Takada T, Shoda J-I, Suzuki H. Novel function of Niemann-Pick C1-like 1 as a negative regulator of Niemann-Pick C2 protein. *Hepatology*. 2012;55:953-64.
45. Yamanashi Y, Takada T, Yoshikado T, Shoda J-I, Suzuki H. NPC2 regulates biliary cholesterol secretion via stimulation of ABCG5/G8-mediated cholesterol transport. *Gastroenterology*. 2011;140:1664-74.
46. Stanley M, Pineda E, Cheng S. Serum cholesterol esters and intestinal cholesterol secretion and absorption in obstructive jaundice due to cancer. *New Engl J Med*. 1959;261:368-73.
47. Cheng S, Stanley M. Secretion of cholesterol by intestinal mucosa in patients with complete common bile duct obstruction. *Proc Soc Exp Biol Med*. 1959;101:223-5.

48. Simmonds WJ, Hofmann AF, Theodor E. Absorption of cholesterol from a micellar solution: intestinal perfusion studies in man. *J Clin Invest.* 1967;46:874-90.
49. Deckelbaum RJ, Lees RS, Small DM, Hedberg SE, Grundy SM. Failure of complete bile diversion and oral bile acid therapy in the treatment of homozygous familial hypercholesterolemia. *New Engl J Med.* 1977;296:465-70.
50. Rosenfeld R, Hellman L. The relation of plasma and biliary cholesterol to bile acid synthesis in man. *J Clin Invest.* 1959;38:1334-8.
51. Hellman L, Rosenfeld R, Eidinoff M, et al. Isotopic studies of plasma cholesterol of endogenous and exogenous origins. *J Clin Invest.* 1955;34:48-60.
52. Stellaard F, Sackmann M, Sauerbruch T, Paumgartner G. Simultaneous determination of cholic acid and chenodeoxycholic acid pool sizes and fractional turnover rates in human serum using ¹³C-labeled bile acids. *J Lipid Res.* 1984;25:1313-9.
53. Jakulj L, Mohammed H, van Dijk TH, et al. Plasma plant sterols serve as poor markers of cholesterol absorption in man. *J Lipid Res.* 2013;54:1144-50.
54. Im S-S, Osborne TF. Liver x receptors in atherosclerosis and inflammation. *Circ Res.* 2011;108:996-1001.
55. Calkin AC, Tontonoz P. Liver x receptor signaling pathways and atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2010;30:1513-8.
56. Bonamassa B, Moschetta A. Atherosclerosis: lessons from LXR and the intestine. *Trends Endocrinol Metab.* 2013;24:120-8.
57. Lo Sasso G, Murzilli S, Salvatore L, et al. Intestinal specific LXR activation stimulates reverse cholesterol transport and protects from atherosclerosis. *Cell Metab.* 2010;12:187-93.
58. Yasuda T, Grillot D, Billheimer JT, et al. Tissue-specific liver X receptor activation promotes macrophage reverse cholesterol transport in vivo. *Arterioscler Thromb Vasc Biol.* 2010;30:781-6.
59. Sudhop T, Lütjohann D, Kodal A, et al. Inhibition of intestinal cholesterol absorption by ezetimibe in humans. *Circulation.* 2002;106:1943-8.
60. Davis HR, Zhu L-J, Hoos LM, et al. Niemann-Pick C1 Like 1 (NPC1L1) is the intestinal phytosterol and cholesterol transporter and a key modulator of whole-body cholesterol homeostasis. *J Biol Chem.* 2004;279:33586-92.
61. Sehayek E, Hazen SL. Cholesterol absorption from the intestine is a major determinant of reverse cholesterol transport from peripheral tissue macrophages. *Arterioscler Thromb Vasc Biol.* 2008;28:1296-7.
62. Briand F, Naik SU, Fuki I, et al. Both the peroxisome proliferator-activated receptor delta agonist, GW0742, and ezetimibe promote reverse cholesterol transport in mice by reducing intestinal reabsorption of HDL-derived cholesterol. *Clin Transl Sci.* 2009;2:127-33.
63. Tang W, Ma Y, Jia L, Ioannou YA, Davies JP, Yu L. Genetic inactivation of NPC1L1 protects against sitosterolemia in mice lacking ABCG5/ABCG8. *J Lipid Res.* 2009;50:293-300.
64. Vrins CLJ, van der Velde AE, van den Oever K, et al. Peroxisome proliferator-activated receptor delta activation leads to increased transintestinal cholesterol efflux. *J Lipid Res.* 2009;50:2046-54.
65. Wang HH, Portincasa P, Mendez-Sanchez N, Uribe M, Wang DQ-H. Effect of ezetimibe on the prevention and dissolution of cholesterol gallstones. *Gastroenterology.* 2008;134:2101-10.
66. Zúñiga S, Molina H, Azocar L, et al. Ezetimibe prevents cholesterol gallstone formation in mice. *Liver Int.* 2008;28:935-47.
67. Davis HR, Lowe RS, Neff DR. Effects of ezetimibe on atherosclerosis in preclinical models. *Atherosclerosis.* 2011;215:266-78.
68. Kastelein JJP, Akdim F, Stroes ESG, et al. Simvastatin with or without ezetimibe in familial hypercholesterolemia. *New Engl J Med.* 2008;358:1431-43.
69. Taylor AJ, Villines TC, Stanek EJ, et al. Extended-release niacin or ezetimibe and carotid intima-media thickness. *New Engl J Med.* 2009;361:2113-22.
70. Bogiatzi C, Spence JD. Ezetimibe and regression of carotid atherosclerosis: importance of measuring plaque burden. *Stroke.* 2012;43:1153-5.
71. Villines TC, Stanek EJ, Devine PJ, et al. The ARBITER 6-HALTS Trial (Arterial Biology for the Investigation of the Treatment Effects of Reducing Cholesterol 6-HDL and LDL Treatment Strategies in Atherosclerosis): final results and the impact of medication adherence, dose, and treatment duration. *J Am Coll Cardiol.* 2010;55:2721-6.
72. Engelking LJ, McFarlane MR, Li CK, Liang G. Blockade of cholesterol absorption by ezetimibe reveals a complex homeostatic network in enterocytes. *J Lipid Res.* 2012;53:1359-68.
73. Descamps OS, De Sutter J, Guillaume M, Missault L. Where does the interplay between cholesterol absorption and synthesis in the context of statin and/or ezetimibe treatment stand today? *Atherosclerosis.* 2011;217:308-21.
74. Califf RM, Lokhnygina Y, Cannon CP, et al. An update on the IMPROVED reduction of outcomes: Vytorin Efficacy International Trial (IMPROVE-IT) design. *Am Heart J.* 2010;159:705-9.
75. Petrides F, Shearston K, Chatelais M, Guilbaud F, Meilhac O, Lambert G. The promises of PCSK9 inhibition. *Curr Opin Lipidol.* 2013;24:307-12.
76. Cohen JC, Boerwinkle E, Mosley TH, Hobbs HH. Sequence variations in PCSK9, low LDL, and protection against coronary heart disease. *New Engl J Med.* 2006;354:1264-72.
77. Levy E, Ben Djoudi Ouadda A, Spahis S, et al. PCSK9 plays a significant role in cholesterol homeostasis and lipid transport in intestinal epithelial cells. *Atherosclerosis.* 2013;227:297-306.
78. Salen G, Ahrens EH, Grundy SM. Metabolism of beta-sitosterol in man. *J Clin Invest.* 1970;49:952-67.
79. O'Neill FH, Sanders TAB, Thompson GR. Comparison of efficacy of plant stanol ester and sterol ester: short-term and longer-term studies. *Am J Cardiol.* 2005;96:29D-36D.
80. McKenney JM, Davidson MH, Jacobson TA, Guyton JR. Final conclusions and recommendations of the National Lipid Association Statin Safety Assessment Task Force. *Am J Cardiol.* 2006;97:89C-94C.
81. Jones PJ, Raeini-Sarjaz M, Ntanos FY, Vanstone CA, Feng JY, Parsons WE. Modulation of plasma lipid levels and cholesterol kinetics by phytosterol versus phytosterol esters. *J Lipid Res.* 2000;41:697-705.
82. Calpe-Berdiel L, Escolà-Gil JC, Blanco-Vaca F. New insights into the molecular actions of plant sterols and stanols in cholesterol metabolism. *Atherosclerosis.* 2009;203:18-31.
83. Racette SB, Lin X, Lefevre M, et al. Dose effects of dietary phytosterols on cholesterol metabolism: a controlled feeding study. *Am J Clin Nutr.* 2010;91:32-8.
84. Kaneko E, Matsuda M, Yamada Y, Tachibana Y, Shimomura I, Makishima M. Induction of intestinal ATP-binding cassette transporters by a phytosterol-derived liver X receptor agonist. *J Biol Chem.* 2003;278:36091-8.
85. Plat J, Mensink RP. Increased intestinal ABCA1 expression contributes to the decrease in cholesterol absorption after plant stanol consumption. *FASEB J.* 2002;16:1248-53.
86. Calpe-Berdiel L, Escolà-Gil JC, Julve J, Zapico-Muñiz E, Canals F, Blanco-Vaca F. Differential intestinal mucosal protein expression in hypercholesterolemic mice fed a phytosterol-enriched diet. *Proteomics.* 2007;7:2659-66.